

Application of Three-Dimensional Culture Conditions to Human Embryonic Stem Cell-Derived Definitive Endoderm Cells Enhances Hepatocyte Differentiation and Functionality

Thamil Selvee Ramasamy, PhD,^{1,*} Jason S.L. Yu, MRes,¹ Clare Selden, PhD,²
Humphery Hodgson, MD,² and Wei Cui, PhD¹

Human embryonic stem cells (hESCs) and induced pluripotent stem cells (iPSCs) provide an unlimited source for the generation of human hepatocytes, owing to their indefinite self-renewal and pluripotent properties. Both hESC-/iPSC-derived hepatocytes hold great promise in treating liver diseases as potential candidates for cell replacement therapies or as an *in vitro* platform to conduct new drug trials. It has been previously demonstrated that the initiation of hESC differentiation in monolayer cultures increases the generation of definitive endoderm (DE) and subsequently of hepatocyte differentiation. However, monolayer culture may hinder the maturation of hESC-derived hepatocytes, since such two-dimensional (2D) conditions do not accurately reflect the complex nature of three-dimensional (3D) hepatocyte specification *in vivo*. Here, we report the sequential application of 2D and 3D culture systems to differentiate hESCs to hepatocytes. Human ESCs were initially differentiated in a monolayer culture to DE cells, which were then inoculated into Algimatrix scaffolds. Treatments of hESC-DE cells with a ROCK inhibitor before and after inoculation dramatically enhanced their survival and the formation of spheroids, which are distinct from HepG2 carcinoma cells. In comparison with monolayer culture alone, sequential 2D and 3D cultures significantly improved hepatocyte differentiation and function. Our results demonstrate that hESC-DE cells can be incorporated into Algimatrix 3D culture systems to enhance hepatocyte differentiation and function.

Introduction

THE LIVER PLAYS an essential role in metabolism, detoxification, and maintenance of homeostasis in the body. Many of these functions are executed by the hepatocytes, the primary cell type constituting the liver. Currently, the only available treatment for the liver failure is liver transplantation, which is severely limited by the shortage of liver donors. Therefore, hepatocyte transplantation has been considered as a putative alternative treatment and has attracted increasing attention, particularly in recent years with the development of human embryonic stem cells (hESCs) and induced pluripotent stem cells (iPSCs), as these cells could potentially provide an unlimited source for the generation of human hepatocytes.^{1,2} In addition to hepatocyte transplantation, stem cell-derived functional human hepatocytes also have the potential to be a valuable cell source for the development of a bioartificial liver^{3,4} as well as for providing an easily accessible *in vitro* model for determining the metabolic and toxicological properties of potential drugs in humans, a key need unmet in current pharmaceutical development.⁵ Unlike

transplantation, in which hepatic progenitor cells can be further differentiated *in vivo*, these later applications require more matured hepatocytes, which are proficient in the functions of adult liver cells, especially in their metabolic functions and toxicological responses to different drug treatments.

Over the last decade, significant progress has been made in developing methods to differentiate pluripotent stem cells efficiently to hepatocyte-like cells (HLCs), through the sequential application of growth factors that are known to play an important role at various stages of liver development.⁶⁻⁹ The resulting HLCs exhibit a typical hepatocyte morphology, express almost all the hepatocyte markers, and perform a wide range of hepatocyte functions. However, the functionality of *in vitro* differentiated hepatocytes is lower than in primary hepatocytes. This is hardly surprising, given that functional performance of primary hepatocytes deteriorates after a short period in culture,¹⁰ indicating that the culture conditions in these experiments are suboptimal for promoting and maintaining hepatocyte functionality. Therefore, there is a need to improve hepatocyte differentiation and culture conditions to enhance their functionality.

¹Department of Surgery and Cancer, Institute of Reproductive and Developmental Biology, Imperial College London, London, United Kingdom.

²Centre for Hepatology, UCL Medical School, University College London, London, United Kingdom.

*Current affiliation: Department of Molecular Medicine, Faculty of Medicine, University of Malaya, Kuala Lumpur, Malaysia.

The majority of studies to date involving hepatocyte differentiation have been carried out in two-dimensional (2D) monolayer culture conditions throughout,^{6,7,11} or have been initiated with embryoid body (EB) formation, followed by 2D cultures.¹² Monolayer culture is thought to have certain advantages over EB formation for the efficient differentiation of pluripotent stem cells to specific lineages, as it enables all the cells to receive a similar level of stimuli from the culture medium and neighboring cells, compared with EB initiation, where cells receive a diverse range of signal inputs depending on their location within the EB. This difference may be especially crucial for the efficient generation of definitive endoderm (DE), since high levels of the growth factor Activin A through supplementation in the culture medium are necessary for the specification of hESCs to DE cells.¹³ However, since liver development is also regulated by cell–cell contact, the absence of suitable cell contacts in the 2D culture conditions may contribute to the reduced functionality. Indeed, it has been demonstrated that a three-dimensional (3D) culture can reduce the hepatocyte functional decline that is seen using the 2D culture system.¹⁴ Therefore, we hypothesized that transference from an initial 2D culture condition to a 3D system would significantly improve hepatocyte maturation and concurrently their functionality. Here, we report the differentiation of hESCs to hepatocytes utilizing a combination of 2D and 3D culture systems. We demonstrate that DE cells derived from monolayer differentiation of hESCs have the ability to form aggregates in Alginmatrix scaffold 3D culture when treated with a Rho-associated kinase (ROCK) inhibitor. Residing in this 3D culture microenvironment therefore enhances subsequent hepatocyte maturation, leading to a clear improvement in their functionality.

Materials and Methods

Culture and differentiation of hESCs

The hESC line H1 was routinely cultured and propagated in Matrigel-coated plates with a mouse embryonic fibroblast-conditioned medium supplemented with basic fibroblast growth factor as previously described.¹⁵ hESCs were differentiated into hepatocytes in monolayer culture using the protocol previously established in our laboratory (Fig. 1A).⁷ Briefly, differentiation was initiated by culturing for 24 h with 100 ng/mL Activin A and 1 mM sodium butyrate (NaB), followed by 2 days with 100 ng/mL Activin A and 0.5 mM NaB. The cells were then cultured for 5 days in a stage II medium containing 20% KO serum replacement and 0.1% dimethyl sulfoxide (DMSO), followed by 4–5 days in stage III medium (L15 medium containing 8.3% fetal bovine serum, 8.3% tryptose phosphate broth, 10 μ M hydrocortisone 21-hemisuccinate, 1 μ M insulin and 2 mM glutamine, 10 ng/mL hepatocyte growth factor, and 20 ng/mL Oncostatin M). For 3D differentiation, hESCs were differentiated in the same medium as in the monolayer cultures until the second day of stage II when they were inoculated into the Alginmatrix plate (Invitrogen 12684-023) (Fig. 1A). Before inoculation, cells were treated with 10 μ M ROCK inhibitor Y-27632 (ROCKi) for 2 h before they were detached from the culture dish with accutase. Viable cells were transferred into a 24-well Alginmatrix plate by centrifugation at 200 g for 3 min at either 0.5×10^6 or 1×10^6 cells per well in a medium also supplemented with ROCKi, which was withdrawn after 48 h.

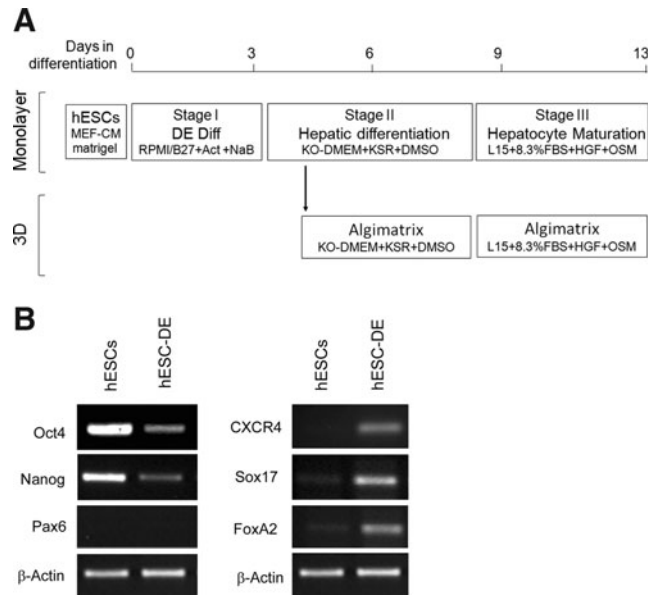


FIG. 1. Differentiation of human embryonic stem cells (hESCs) to hepatocyte-like cells by sequential application of two-dimensional (2D) and three-dimensional (3D) culture systems. **(A)** Schematic illustration of the procedure to differentiate hESCs to hepatocytes. Human ESCs were initially differentiated in a monolayer culture until day 4 (2nd day in the stage II medium), and then the cells were transferred into Alginmatrix 3D culture system for further differentiation to hepatocytes. Act, Activin A; FBS, fetal bovine serum; HGF, hepatocyte growth factor; NaB, sodium butyrate; OSM, Oncostatin M. **(B)** Semiquantitative reverse transcription–polymerase chain reaction (RT-PCR) analysis showing gene expression in hESCs and their differentiated definitive endoderm (hESC-DE) cells before transfer to the 3D culture. Pluripotent and neural genes are on the left panel, and definitive endoderm genes are on the right panel.

Analysis of the spheroids

Cell spheroids were released from the scaffold by 5-min incubation with 1 mL dissolving buffer (Invitrogen A11340-01) and were then harvested by centrifugation at 200 g for 4 min. Spheroids were stained by 90 s of incubation with propidium iodide (PI) resuspended at 40 μ g/mL in phosphate-buffered saline (PBS), and images were captured by a Nikon E2000 phase-contrast fluorescent microscope. Dimensional analysis of the spheroids was performed using images taken under a phase-contrast microscope and using the medium axis of the spheroid. To determine the viable cell number, spheroids were dissociated into single cells by 8–10-min incubation with trypsin–ethylenediaminetetraacetic acid at 37°C, during which time the spheroids were gently resuspended twice using a pipette. Single cells were then stained with trypan blue and counted with a hemocytometer.

RNA extraction and reverse transcription–polymerase chain reaction

Total RNA was extracted using TRI reagent solution (Sigma T9424) following the manufacturer's instructions. Remaining traces of DNAs were removed by DNase I treatment. First-strand cDNA was synthesized from 2 μ g

then inoculated into a 24-well Alginmatrix 3D culture plate at $0.5\text{--}1 \times 10^6$ cells per well. HepG2 cells were also inoculated into the plate in the same way as hESC-DE cells. Interestingly, spheroids were observed in HepG2 cells after 5 days in the 3D culture, but none were visible in hESC-DE cultures (Fig. 2A). PI staining revealed considerable cell death in the hESC-DE cultures (Fig 2B-upper panel). As the HepG2 cells grew well in the 3D culture plate, we surmised that the scaffold itself does not induce any detrimental effect on the cells that leads to their death. As such, we reasoned that dissociation of the hESC-DE cells might be the cause of the high levels of cell death observed postinoculation. It has been reported previously that dissociation of hESCs into single cells increases ROCK-dependent hyperactivation of actin-myosin contraction, which induces apoptosis,^{16,17} and that treatment of hESCs with the ROCK inhibitor Y-27632 (ROCKi) significantly reduces apoptosis after dissociation.¹⁸ To test whether use of ROCK inhibitor could reduce hESC-DE cell death after dissociation, the cells were pretreated with $10\ \mu\text{M}$ ROCKi for 2 h before disassociation, and the treatment was continued for further 48 h after inoculation.

With this treatment, spheroids were clearly observed 24 h after inoculation and were well formed after 9 days with no signs of detectable apoptosis (Fig. 2B). By contrast, ROCKi treatment did not show a significant effect on their ability to form spheroids in HepG2 cells. These results therefore suggest that hESC-DE cells are different from HepG2 cells and require the inhibition of the ROCK signaling pathway to survive and to form 3D structures in Alginmatrix scaffold.

hESC-DE cells form spheroid in the 3D culture via cell aggregation

Since different growth behaviors were observed with hESC-DE cells and HepG2 cells upon their inoculation into the Alginmatrix scaffold, we next examined how spheroids are formed in these two cell types, by monitoring the cells immediately after the inoculation. To our surprise, compact 3D aggregates were already visible in hESC-DE cells 3 h after inoculation, with larger aggregates formed when more cells were inoculated, whereas HepG2 cells still appeared as single cells or loose-contacted cell clusters at the time (Fig. 3A,

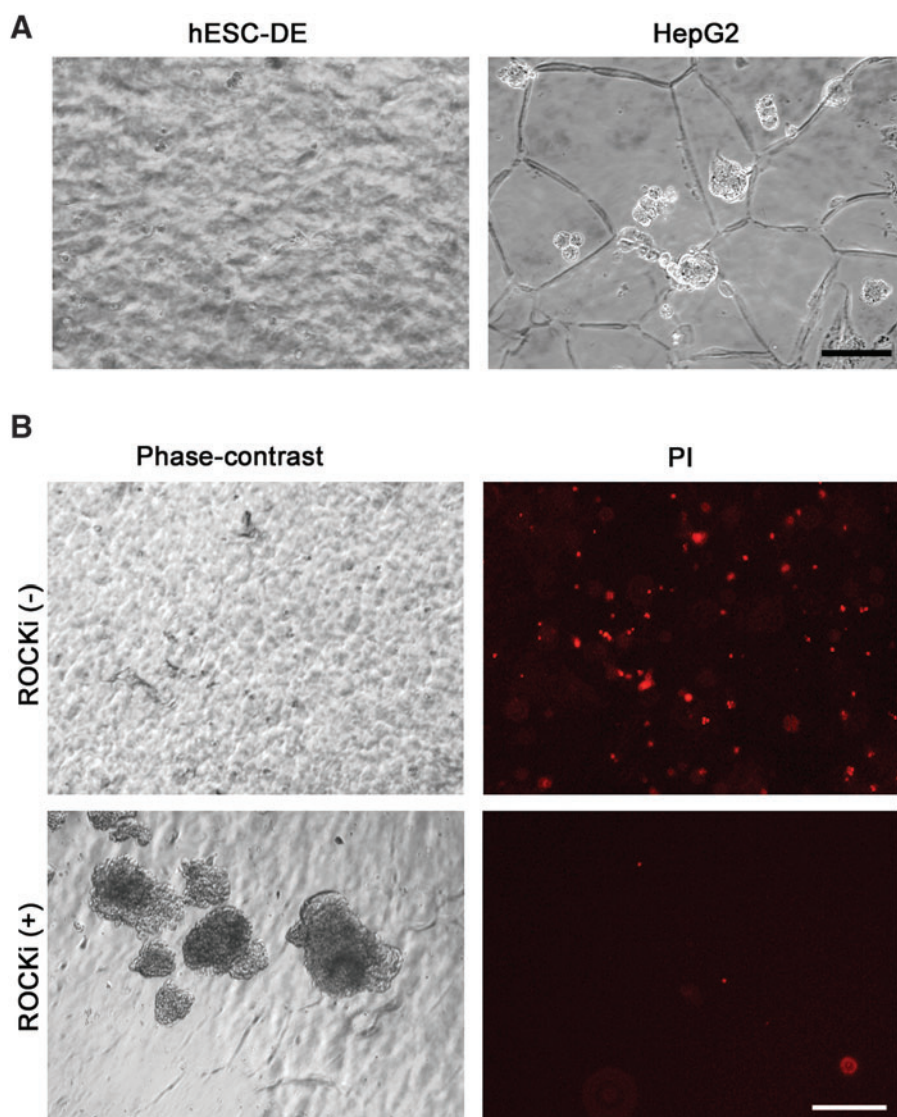
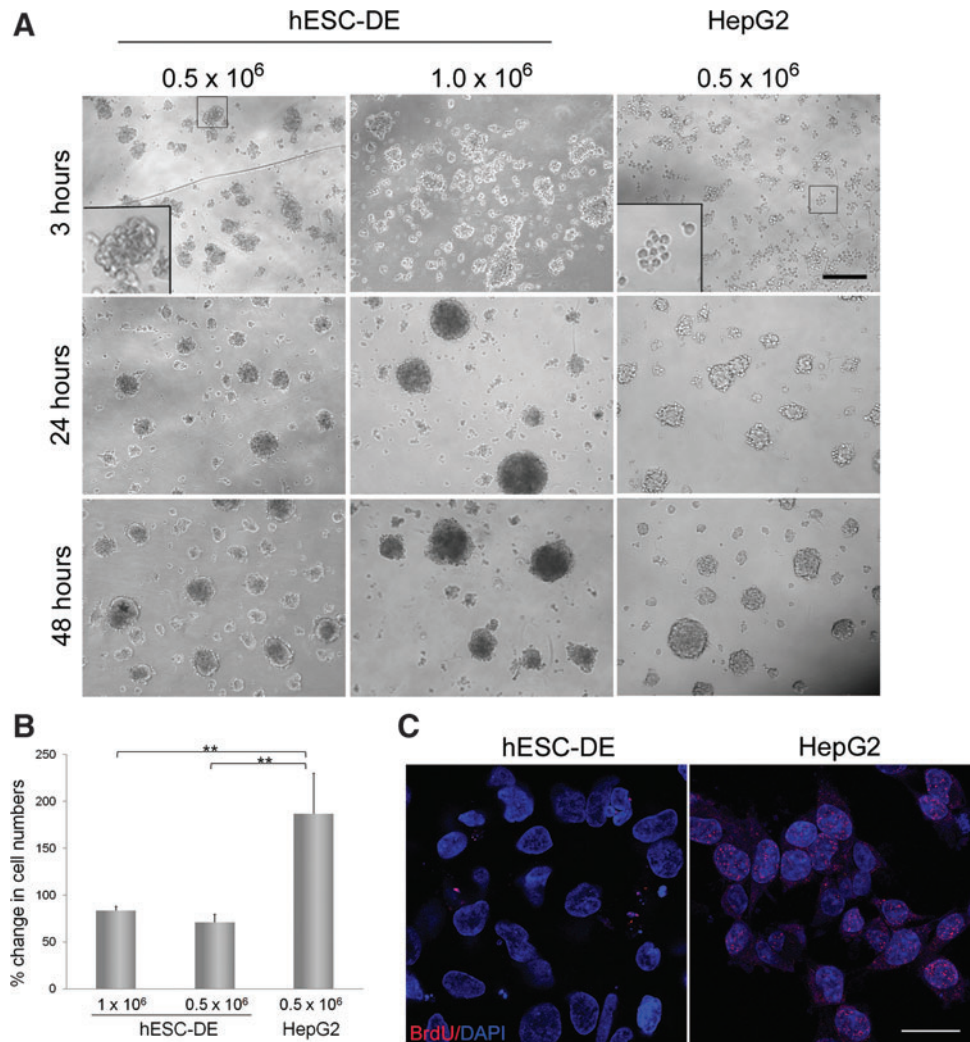


FIG. 2. Effect of ROCK inhibitor (ROCKi) on cell survival and spheroid formation of hESC-DE cells in Alginmatrix scaffold. (A) Both hESC-DE and HepG2 cells were inoculated into 24-well Alginmatrix at 5×10^5 cells/well. Images were taken 5 days after inoculation, showing no spheroids formed in hESC-DE cultures, but evidence of spheroids in HepG2 cells. (B) hESC-DE cells were treated with or without ROCKi before and 48h after inoculation of 1×10^6 cells/well into the Alginmatrix 3D culture. Cells were collected 9 days after the inoculation and stained with propidium iodide (PI) to detect any cell death. ROCKi treatment dramatically reduced the cell death and enhanced the spheroid formation. Scale bar = $100\ \mu\text{m}$. Color images available online at www.liebertpub.com/tea

FIG. 3. Spheroid formation and cell proliferation in Algimatrix 3D cultures. **(A)** Both hESC-DE and HepG2 cells aggregate to form spheroids during the first day after inoculation into the Algimatrix scaffold with hESC-DE cells forming aggregates quicker than the HepG2 cells and higher seeding density producing larger spheroids. In addition, spheroids formed by hESC-DE cells exhibit no significant difference in size between 24–48 h, while spheroids of HepG2 cells show clear increase during this time period. Scale bar = 100 μm . **(B)** Changes in cell number after 9 days in 3D culture. Cell counts are presented as percentage of the initial seeded cell numbers. Standard error bars were calculated from six independent experiments. $**p < 0.001$ (Student's *t*-tests). **(C)** BrdU-labeling assay. Spheroids were incubated with BrdU for 48 h and then dissociated onto a coverslip for further 24 h before stained with an antibody against BrdU. Scale bar = 25 μm . Color images available online at www.liebertpub.com/tea



upper panel). By 24 h in the 3D scaffold, hESC-DE aggregates appeared as spheroids, and similar spheroids were also apparent in HepG2 cells (Fig. 3A, middle panel). The size of spheroids in the hESC-DE cells did not change significantly, whereas the spheroids in the HepG2 cells gradually increased in size during the first 48 h in 3D culture (Fig. 3A, lower panel). These data suggest that hESC-DE cells may form spheroids predominantly via cell aggregation, which can occur within a relatively short time period and may be promoted by a ROCK inhibitor, whereas HepG2 cells may form spheroids through cell aggregation as well as cell proliferation.

To validate this observation, we compared the total viable cell numbers seeded at inoculation with those after 9 days of 3D culture in both cell types. As expected, the number of viable HepG2 cells in the 3D culture notably increased during this time period, although not to the same extent as in the 2D culture, whereas the number of hESC-DE cells showed a decrease in numbers, most likely due to cell death (Fig. 3B). To further confirm the proliferative properties of the two cell types, we incubated the cells with BrdU for 48 h after 3D inoculation, and then stained the cells with an anti-BrdU antibody. HepG2 cells showed positive signals with the BrdU antibody, whereas positive staining was not detected

in the nuclei of hESC-DE cells. Therefore, these results confirm that hESC-DE cells do not proliferate after inoculation into the Algimatrix scaffold.

Cell density affects the size of spheroids in the Algimatrix scaffold

Spheroid size has been reported to affect hepatocyte viability and function in the 3D culture.¹⁴ Since the cell spheroids in culture do not have a vascular network, cells constituting the center of the spheroids become hypoxic and have a reduced uptake of oxygen from the cell medium if they are larger than 100 μm in size. On the other hand, if the spheroid size is too small, they produce significantly less albumin as a result of poor maturation.¹⁹ To study factors that affect the spheroid size, we categorized the spheroids produced by our 3D culture into three groups according to their size: small with spheroids of <50 μm in diameter, medium between 50 and 100 μm in diameter, and larger over 100 μm in diameter, and examined the effect of initial inoculation seeding density on the spheroid size. We measured and categorized ~ 100 spheroids from each inoculation condition 9 days after the cells were inoculated. The results show that in hESC-DE cells, a majority (84%) of spheroids

produced from the lower seeding density of 5×10^5 cells/well fell into the medium-size bracket, with <10% of spheroids falling into the larger- or smaller-size group. However, at higher seeding density of 1×10^6 cells/well, almost 20% and 30% of the spheroids were in the large- and smaller-size group, respectively, with <60% in the medium-size category (Fig. 4). In HepG2 cells, the lower seeding density of 5×10^5 cells/well produced very few spheroids (<5%) of the large size. These results suggest that the initial seeding density of hESC-DE cells has an effect on the sizes of the spheroids formed in the 3D scaffold.

Algimatrix 3D culture promotes hepatocyte differentiation and function

Alginate scaffold has been found to promote cells from the liver of newborn rats to differentiate into functional liver tissues.²⁰ To evaluate whether the Algimatrix 3D culture has any advantage over monolayer culture in differentiating DE cells into hepatocytes, we compared the hepatic gene expression profiles in the 3D culture to those in monolayer as well as to undifferentiated hESCs and the hESC-DE cells used at the 3D inoculation stage. Unsurprisingly, undifferentiated hESCs exhibited high levels of the pluripotent gene *Oct4*, but showed no expression of hepatic markers. Expression of the DE marker, *Sox17*, was apparent in hESC-DE cells, but neither *Oct4* nor any hepatic genes were detectable. In contrast, hESC-DE cells that had been further differentiated in both 2D and 3D culture conditions expressed the hepatocyte genes, albumin, apolipoprotein F (ApoF), tryptophan dioxygenase (TO), and cytochrome P450 family members, CYP3A4 and CYP7A1 (Fig. 5A). Significantly, the cells differentiated in the Algimatrix 3D scaffold exhibited considerably higher levels of hepatocyte-specific markers than the cells differentiated in monolayer, particularly those that are expressed predominantly in the more mature hepatocytes, such as ApoF, TO, and CYP3A4.²¹

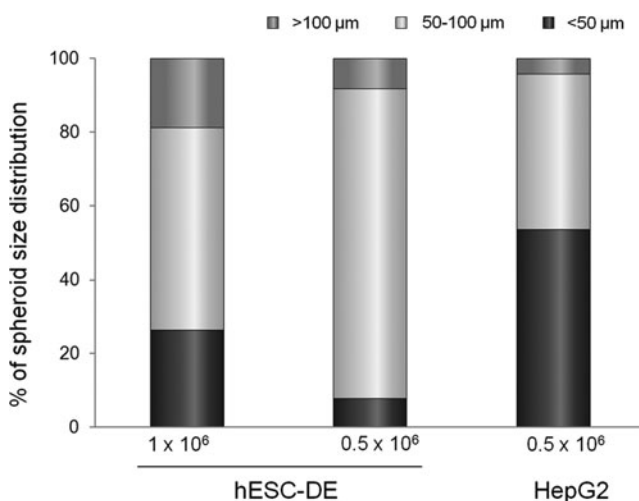


FIG. 4. Effect of seeding density on spheroids' sizes. Spheroids from each culture were released after 9 days in the Algimatrix scaffold, and their size was measured from images taken with a phase-contrast microscope as described in the Materials and Methods section. At least 100 randomly selected spheroids were measured from each culture condition and grouped according to their size.

Since CYP3A4 is the most crucial P450 enzymes in the human liver, metabolizing about half of all drugs on the market today, we examined CYP3A4 activity in hESC-DE and HepG2 cells that were either cultured in 3D conditions or differentiated in monolayer to further determine the improvement of 3D culture on the differentiation and function of hepatocytes. In comparison with the monolayer culture system, at the lower seeding density of 0.5×10^6 cells/well, Algimatrix 3D culture significantly increased basal levels of CYP3A4 activity in both hESC-DE cells and HepG2 cells (Fig. 5B). The higher inoculation density also increased activity, but to a lesser extent, probably due to a suboptimal spheroid size. Furthermore, the cells cultured in the 3D condition also exhibited increased CYP3A4 activity in response to rifampicin induction (Fig. 5C). To further validate the improvement of 3D culture on hepatocyte differentiation and function, we measured urea secretion in hESC-derived hepatocytes from both 2D and 3D cultures (Fig. 5D), which showed that Algimatrix 3D culture significantly enhanced urea secretion when hESC-DE cells were seeded at 0.5×10^6 cells/well. Taken together, we have shown that Algimatrix 3D culture system improves the differentiation of hESC-DE cells to hepatocytes, and as a result, enhances the expression of hepatocyte markers and certain functions.

Discussion

We have demonstrated here that DE cells derived from hESCs are able to form spheroids in an Algimatrix scaffold. In comparison with the monolayer culture, Algimatrix 3D culture enhances hepatocyte differentiation and function. These results correspond with the previous findings from hESCs.⁹ However, in the previous studies, the 3D culture was directly applied to the hESCs, in which hESCs were detached into small clumps and cultured in a low-adherent plate to form EBs. Since pluripotent hESCs in the EBs receive variable signals depending on their position, this method may have an increased chance of inducing the differentiation of cells to other lineages, which consequently reduces the final yield of hepatocytes. Our previous studies, as well as those of others, have shown that monolayer culture supplemented with high-dosage Activin A can differentiate hESCs efficiently to the DE cells that are a prerequisite for the generation of hepatocytes.^{7,13} Therefore, to increase hepatocyte production, we incorporated the 3D culture at a later stage in the differentiation protocol, once the hESCs were committed to DE. We differentiated hESCs for 4 days in monolayer culture with our established method and demonstrated that the cells were primarily differentiated into DE (Fig. 1B) before incorporating them into 3D culture.

Here, and in accordance with others, we have demonstrated that an alginate-based 3D culture system can enhance the functionality of HepG2 cells and has no toxic effects on the cells.^{22,23} Interestingly, we have also shown that hESC-DE cells behave differently from HepG2 cells when cultured in an Algimatrix 3D scaffold. Firstly, hESC-DE cells required the treatment of ROCKi to survive in the 3D culture after dissociation, whereas HepG2 cells do not require ROCKi to survive or to form spheroids. In addition, treatment of HepG2 cells with ROCKi showed no effect on their survival and proliferation. This phenotype of hESC-DE cells is similar to that reported in undifferentiated hESCs.²⁴ In hESCs,

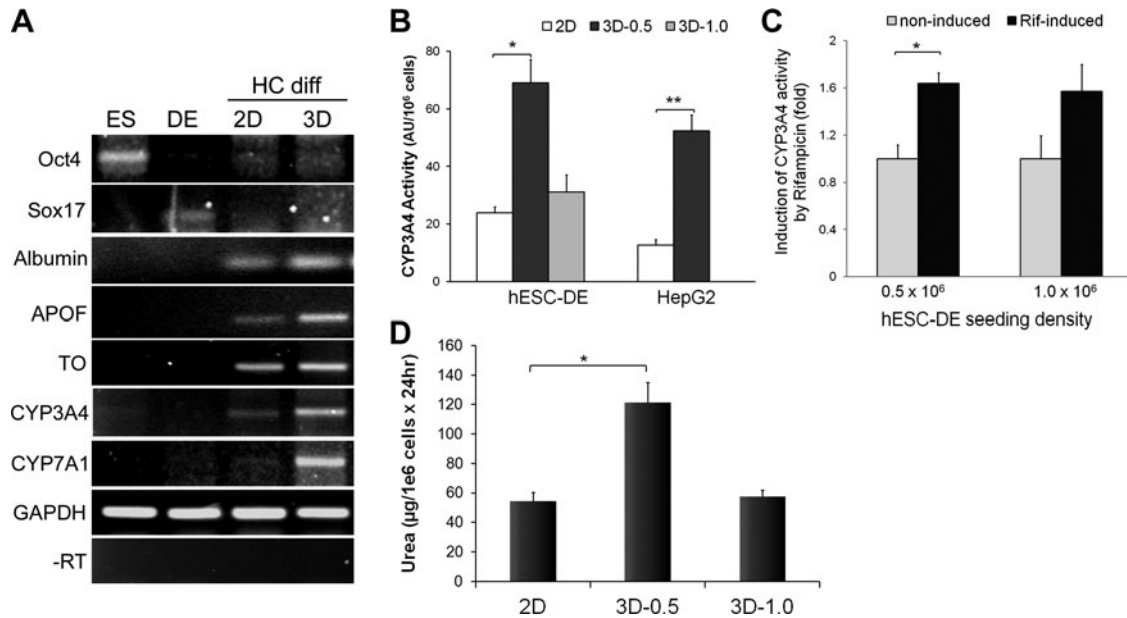


FIG. 5. Comparison of the effect of 2D (monolayer) and 3D culture conditions on hepatocyte marker expression and functionality. **(A)** Semiquantitative RT-PCR analysis on gene expression in undifferentiated hESCs (ES), hESC-DE cells before 3D culture (DE), and hepatocytes differentiated via 2D and 3D culture conditions as indicated on Fig 1A. APOF, apolipoprotein F; TO, tryptophan dioxygenase. Total RNAs without RT exhibited no signals with all primers. **(B)** Algimatrix 3D cultures enhance CYP3A4 activity in both hESC-DE and HepG2 cells. Cells were seeded into the 3D Algimatrix plate at 0.5×10^6 (3D-0.5) or 1×10^6 (3D-1) cells per well and differentiated for 9 days, CYP3A4 activity was then measured as described in the Materials and Methods section. Error bars were calculated from three independent experiments. $*p < 0.01$ and $**p < 0.005$ (Student's *t*-test). **(C)** Increase of CYP3A4 activity in hESC-DE-derived hepatocytes in response to rifampicin treatments. $*p < 0.05$. **(D)** Urea secretion in hESC-DE-derived hepatocytes from 2D and Algimatrix 3D cultures seeded at 0.5×10^6 (3D-0.5) or 1×10^6 (3D-1) cells per well. Error bars were calculated from three independent experiments. $*p < 0.01$.

apoptosis upon dissociation is thought to be caused by ROCK-dependent actomyosin hyperactivation, which is triggered by the loss of E-cadherin-dependent intercellular contact.¹⁷ However, hESC-DE cells stained with anti-E-cadherin antibody exhibited no clear positive signals (data not shown), indicating that the apoptosis induced by dissociation in hESC-DE cells is not a downstream effect of E-cadherin. Nonetheless, one cannot exclude the possibility that this phenotype could still be related to the loss of attachment-dependent actin-myosin contraction.¹⁶ Further experiments are required to elucidate the underlying mechanisms. Secondly, hESC-DE cells aggregate more rapidly than the HepG2 cells. Thirdly, unlike HepG2 cells, hESC-DE cells were unable to proliferate in the Algimatrix 3D scaffold, which was demonstrated by the fact that there was no significant increase in the cell numbers after 9 days in the 3D culture, and that BrdU labeling showed no incorporation (Fig. 3). This was also different from their behavior in monolayer culture, in which hESC-DE cells were able to divide and incorporate BrdU. Although the exact mechanisms remain unclear, it has been demonstrated previously that the growth properties and differentiated functionality of rat hepatocytes are reciprocally modulated in culture by cell-cell contact, and that this reciprocal modulation is lost in hepatoma cells.^{25,26} Therefore, it is likely that increased cell-cell contact may account for the phenotype, which may also contribute to the improved differentiation and functionality.

We have analyzed spheroid formation and hepatocyte function of hESC-DE cells at two different seeding densities. The two different seeding densities did not seem to have a

clear effect on the timing of spheroid formation, but did appear to impact the size of spheroids formed in the Algimatrix scaffold. However, this effect was not a simple positive correlation. Surprisingly, the higher seeding density not only resulted in a higher proportion of the larger spheroids but also increased the number of smaller spheroids. In addition, our results showed that lower seeding densities that generated significantly more medium-sized spheroids had higher levels of CYP3A4 activity and a significant induction upon drug stimulation as well as increased urea secretion, indicating that the size of hESC-DE spheroids is important for the hepatocyte differentiation and function. It is not surprising that it is the medium-sized spheroids that exhibit better functionality, as the larger size reduces the nutrition/oxygen accessibility of the cells found at the center and affects the survival of these cells; while the smaller spheroids do not form the proper cell-cell contacts, which may subsequently hinder hepatocyte differentiation and function.¹⁹ In summary, our study has demonstrated that hESC-DE cells can be incorporated into an Algimatrix scaffold 3D culture upon treatment with the ROCK inhibitor. The 3D culture enhances hESC-DE cell differentiation to HLCs and improves the resulting hepatocyte function, such as CYP3A4 activity.

Acknowledgments

We thank Dr. Sam Coward and Dr. Isobel Massie for their technical help and discussion at early stage of the project and thank Dr. Fiona Lamont for their helpful discussion on the

manuscript. T.S.R. was supported by a studentship from the University of Malaya/Ministry of Higher Education, Malaysia. This work is supported by the Genesis Research Trust.

Disclosure Statement

No competing financial interests exist.

References

1. Irion, S., Nostro, M.C., Kattman, S.J., and Keller, G.M. Directed differentiation of pluripotent stem cells: from developmental biology to therapeutic applications. *Cold Spring Harb Symp Quant Biol* **73**, 101, 2008.
2. Stutchfield, B.M., Forbes, S.J., and Wigmore, S.J. Prospects for stem cell transplantation in the treatment of hepatic disease. *Liver Transpl* **16**, 827, 2010.
3. Selden, C., and Hodgson, H. Cellular therapies for liver replacement. *Transpl Immunol* **12**, 273, 2004.
4. Behbahan, I.S., Duan, Y., Lam, A., Khoobyari, S., Ma, X., Ahuja, T.P., and Zern, M.A. New approaches in the differentiation of human embryonic stem cells and induced pluripotent stem cells toward hepatocytes. *Stem Cell Rev* **7**, 748, 2011.
5. Guguen-Guillouzo, C., Corlu, A., and Guillouzo, A. Stem cell-derived hepatocytes and their use in toxicology. *Toxicology* **270**, 3, 2010.
6. Cai, J., Zhao, Y., Liu, Y., Ye, F., Song, Z., Qin, H., Meng, S., Chen, Y., Zhou, R., Song, X., *et al.* Directed differentiation of human embryonic stem cells into functional hepatic cells. *Hepatology* **45**, 1229, 2007.
7. Hay, D.C., Zhao, D., Fletcher, J., Hewitt, Z.A., McLean, D., Urruticoechea-Uriguen, A., Black, J.R., Elcombe, C., Ross, J.A., Wolf, R., *et al.* Efficient differentiation of hepatocytes from human embryonic stem cells exhibiting markers recapitulating liver development *in vivo*. *Stem Cells* **26**, 894, 2008.
8. Green, M.D., Chen, A., Nostro, M.C., d'Souza, S.L., Schaniel, C., Lemischka, I.R., Gouon-Evans, V., Keller, G., and Snoeck, H.W. Generation of anterior foregut endoderm from human embryonic and induced pluripotent stem cells. *Nat Biotechnol* **29**, 267, 2011.
9. Baharvand, H., Hashemi, S.M., Kazemi, A.S., and Farrokhi, A. Differentiation of human embryonic stem cells into hepatocytes in 2D and 3D culture systems *in vitro*. *Int J Dev Biol* **50**, 645, 2006.
10. Elaut, G., Henkens, T., Papeleu, P., Snykers, S., Vinken, M., Vanhaecke, T., and Rogiers, V. Molecular mechanisms underlying the dedifferentiation process of isolated hepatocytes and their cultures. *Curr Drug Metab* **7**, 629, 2006.
11. Bone, H.K., Nelson, A.S., Goldring, C.E., Tosh, D., and Welham, M.J. A novel chemically directed route for the generation of definitive endoderm from human embryonic stem cells based on inhibition of GSK-3. *J Cell Sci* **124**, 1992, 2011.
12. Lavon, N., Yanuka, O., and Benvenisty, N. Differentiation and isolation of hepatic-like cells from human embryonic stem cells. *Differentiation* **72**, 230, 2004.
13. D'Amour, K.A., Agulnick, A.D., Eliazar, S., Kelly, O.G., Kroon, E., and Baetge, E.E. Efficient differentiation of human embryonic stem cells to definitive endoderm. *Nat Biotechnol* **23**, 1534, 2005.
14. Glicklis, R., Shapiro, L., Agbaria, R., Merchuk, J.C., and Cohen, S. Hepatocyte behavior within three-dimensional porous alginate scaffolds. *Biotechnol Bioeng* **67**, 344, 2000.
15. Gerrard, L., Rodgers, L., and Cui, W. Differentiation of human embryonic stem cells to neural lineages in adherent culture by blocking bone morphogenetic protein signaling. *Stem Cells* **23**, 1234, 2005.
16. Chen, G., Hou, Z., Gulbranson, D.R., and Thomson, J.A. Actin-myosin contractility is responsible for the reduced viability of dissociated human embryonic stem cells. *Cell Stem Cell* **7**, 240, 2010.
17. Ohgushi, M., Matsumura, M., Eiraku, M., Murakami, K., Aramaki, T., Nishiyama, A., Muguruma, K., Nakano, T., Suga, H., Ueno, M., *et al.* Molecular pathway and cell state responsible for dissociation-induced apoptosis in human pluripotent stem cells. *Cell Stem Cell* **7**, 225, 2010.
18. Watanabe, K., Ueno, M., Kamiya, D., Nishiyama, A., Matsumura, M., Wataya, T., Takahashi, J.B., Nishikawa, S., Nishikawa, S., Muguruma, K., *et al.* A ROCK inhibitor permits survival of dissociated human embryonic stem cells. *Nat Biotechnol* **25**, 681, 2007.
19. Glicklis, R., Merchuk, J.C., and Cohen, S. Modeling mass transfer in hepatocyte spheroids via cell viability, spheroid size, and hepatocellular functions. *Biotechnol Bioeng* **86**, 672, 2004.
20. Dvir-Ginzberg, M., Elkayam, T., and Cohen, S. Induced differentiation and maturation of newborn liver cells into functional hepatic tissue in macroporous alginate scaffolds. *FASEB J* **22**, 1440, 2008.
21. Snykers, S., De, K.J., Rogiers, V., and Vanhaecke, T. *In vitro* differentiation of embryonic and adult stem cells into hepatocytes: state of the art. *Stem Cells* **27**, 577, 2009.
22. Coward, S.M., Legallais, C., David, B., Thomas, M., Foo, Y., Mavri-Damelin, D., Hodgson, H.J., and Selden, C. Alginate-encapsulated HepG2 cells in a fluidized bed bioreactor maintain function in human liver failure plasma. *Artif Organs* **33**, 1117, 2009.
23. Elkayam, T., Amitay-Shaprut, S., Dvir-Ginzberg, M., Harel, T., and Cohen, S. Enhancing the drug metabolism activities of C3A—a human hepatocyte cell line—by tissue engineering within alginate scaffolds. *Tissue Eng* **12**, 1357, 2006.
24. Harb, N., Archer, T.K., and Sato, N. The Rho-Rock-Myosin signaling axis determines cell-cell integrity of self-renewing pluripotent stem cells. *PLoS One* **3**, e3001, 2008.
25. Nakamura, T., Yoshimoto, K., Nakayama, Y., Tomita, Y., and Ichihara, A. Reciprocal modulation of growth and differentiated functions of mature rat hepatocytes in primary culture by cell—cell contact and cell membranes. *Proc Natl Acad Sci U S A* **80**, 7229, 1983.
26. Nakamura, T., Nakayama, Y., Teramoto, H., Nawa, K., and Ichihara, A. Loss of reciprocal modulations of growth and liver function of hepatoma cells in culture by contact with cells or cell membranes. *Proc Natl Acad Sci U S A* **81**, 6398, 1984.

Address correspondence to:

Wei Cui, PhD

Department of Surgery and Cancer

Institute of Reproductive and Developmental Biology

Imperial College London

Hammersmith Campus

Du Cane Road

London W12 0NN

United Kingdom

E-mail: wei.cui@imperial.ac.uk

Received: March 23, 2012

Accepted: August 13, 2012

Online Publication Date: September 21, 2012